

Synthesis of 9-Membered Masked Eneidyne Analogues Possessing DNA Intercalator and Sugar Moieties

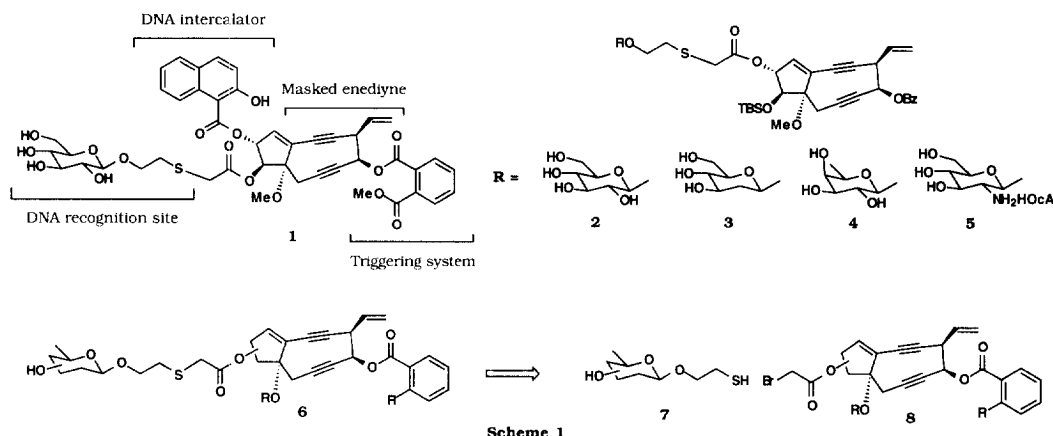
Takashi Takahashi,* Hiroshi Tanaka, Akihisa Matsuda,
Takayuki Doi, and Haruo Yamada

*Department of Chemical Engineering, Tokyo Institute of Technology,
Ookayama, Meguro, Tokyo 152-8552, Japan*

Received 28 August 1998; accepted 6 October 1998

Abstract: Syntheses of hybrid DNA cleaving molecules which have the 9-membered eneidyne structure of kedarcidin with the DNA intercalator of NCS and/or the sugar moieties were accomplished. Introduction of the sugar moieties to the masked eneidyne was achieved via S-alkylation of a bromoacetate derivative with a sugar-containing thiol without the need for protection of the sugar moiety. © 1998 Elsevier Science Ltd. All rights reserved.

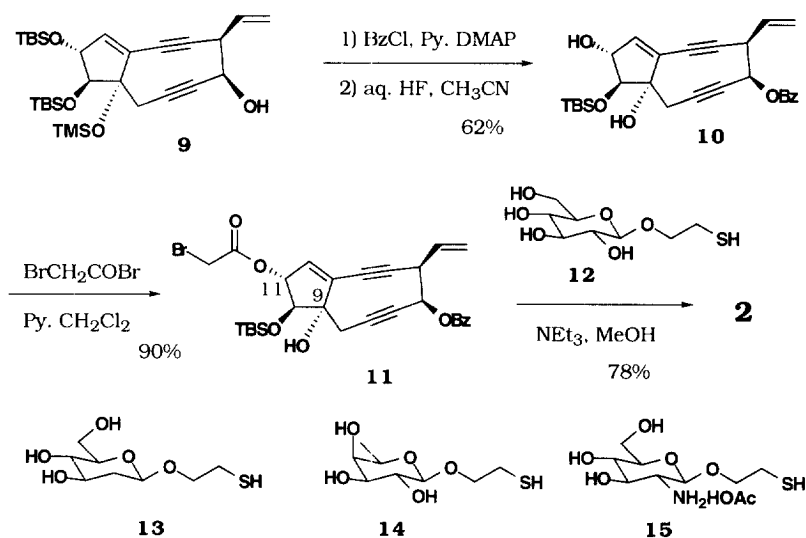
It has been proposed that the eneidyne antitumor antibiotics including neocarzinostatin- (NCS-chr.), C-1027-, kedarcidin-chromophores, and calicheamicin, exert their biological activities through the cycloaromatization of the eneidyne moieties.¹ This aromatization generates a highly reactive biradical species which abstracts hydrogen atoms from deoxyribose resulting in DNA strand scission. Despite the extensive structural diversity of these compounds, they all contain three important functional domains; a) the highly strained eneidyne structure responsible for biradical generation, b) a triggering system to activate cycloaromatization, c) and minor groove binding functionalities, intercalation moieties, and/or combination of these motifs which can assist in association with DNA and in positioning the eneidyne cores for maximum effectiveness. Therefore, hybridization of the 9-membered masked eneidyne structure with a DNA intercalator and/or sugar moiety would be expected to produce new artificial DNA cleaving molecules that have both high DNA cleaving activity and base-sequence specificities. A number of hybrid molecules possessing a DNA cleaving moiety,² triggering system³ and/or DNA binder⁴ have been reported to improve DNA cleaving activity.⁵ In this communication, we describe the synthesis of the masked eneinyne **1**, **2**, **3**, **4** and **5** conjugated with a sugar moiety and/or naphthoate moiety.



Scheme 1

We have previously reported a facile approach to the 9-membered enediynes as analogues of esperamicin-calicheamicin⁶ and NCS-chr.⁷ This concept was applied to the synthesis of analogues **1**, **2**, **3**, **4** and **5**. Our synthetic plan for the introduction of the sugar moiety to the masked enediyne involves S-alkylation of α bromoacetate derivative **8** with the sugar-containing thiol **7**. (Scheme 1) This method would allow us to couple the labile diyne chemoselectively with the thiol derivative of various sugars possessing hydroxyl and/or amino groups without the need for protections on the sugar moiety.

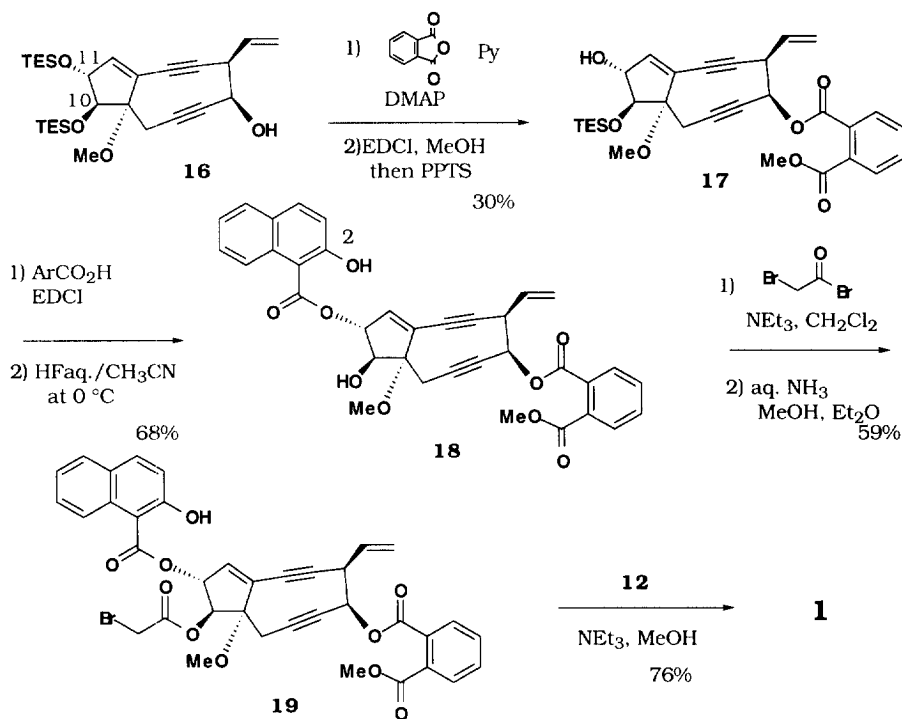
In order to investigate the feasibility of this method, synthesis of diyne **2** conjugated with glucose **12** via a thiol tether at C(11) was examined. (Scheme 2) Benzoylation of **9**, which was prepared according to our previous report,¹¹ followed by selective removal of the TBS group at C(11) and the TMS group at C(9) with aq. HF in CH₃CN gave allylic alcohol **10** in 62% yield. Bromoacetylation of **10** was performed with bromoacetyl bromide in CH₂Cl₂ to afford the desired product **11** in 90% yield. Coupling of **11** with **12** was accomplished by treatment with Et₃N in MeOH at 0 °C for 5 min to provide **2** in 78% yield. Similarly, S-alkylation of bromoacetate **11** with 2-deoxyglucose **13**, fucose **14** and glucosamine **15** afforded the masked analogues **3**, **4** and **5** in 63%, 61% and 59% yields, respectively. The S-alkylation provides a versatile method for introducing the various sugars to the labile enediyne without protecting groups in the sugar moiety.



Scheme 2

Our attention next turned to the diyne **1** possessing naphthoate and glucose moieties. Synthesis of **1** was carried out from the 9-membered diyne **16** which was prepared according to the modified procedure developed in our laboratory.⁷ (Scheme 3) Treatment of the alcohol **16** with phthalic anhydride at room temperature, followed by esterification with 1-ethyl-3-(3-dimethylaminopropyl)-carbodiimide (EDCI) in MeOH gave the phthalate, which was converted to the allylic alcohol **17** by selective removal of the TES group (PPTS, MeOH) at C(11) in 30% overall yield. Naphthoate derivative **18** was prepared by treatment of 2-hydroxy-1-naphthoic acid with EDCI in CH₂Cl₂ and deprotection of the TES group at C(10) with aq. HF

in CH₃CN at 0 °C (68% yield). Treatment of **18** with bromoacetyl bromide in the presence of Et₃N at 0 °C gave the dibromoacetate derivative. Selective removal of the bromoacetyl group at the naphthoate 2-position with aq. ammonia in MeOH afforded bromoacetate **19** in 59% overall yield from **18**. The resulting **19** was coupled with thiol **12** at 0 °C to form the desired analogue **1** in 76% yield after gel permeation chromatography (JAIGEL-1H).



Scheme 3

In summary, we have succeeded in the synthesis of 9-membered masked enediyne analogues possessing a naphthoic acid intercalator and/or various monosaccharides appended via a thiol tether.

Acknowledgment. This work was supported by a Grant-in-Aid for Scientific Research on Priority Area No. 06240104 from the Ministry of Education, Science, Sports and Culture, Japan.

References and Notes

- Reviews: a) Nicolaou, K. C.; Smith, A. L.; Yue, E. W. *Proc. Natl. Acad. Sci. U.S.A.* **1993**, *90*, 5881. b) Maier, M. *Synlett* **1995**, 13. c) Nicolaou, K. C.; Dai, W.-M. *Angew. Chem. Int. Ed. Engl.* **1991**, *30*, 1387. d) Nicolaou, K. C.; Smith, A. L. *Acc. Chem. Res.* **1992**, *25*, 497.
- a) Wender, P. A.; Harmata, M.; Jeffrey, D.; Mukai, C.; Suffert, J. *Tetrahedron Lett.* **1988**, *29*, 909. b) Wender, P. A.; McKinney, J. A.; Mukai, C. *J. Am. Chem. Soc.* **1990**, *112*, 5369. c) Magnus, P.; Pitterna, T. *J. Chem. Soc., Chem. Commun.* **1991**, 541. d) Magnus, P.; Davies, M. *J. Chem. Soc., Chem. Commun.* **1991**, 1522. e) Myers, A. G.; Harrington, P. M.; Kuo, E. Y. *J. Am. Chem. Soc.* **1991**, *113*, 694. f) Myers, A. G.; Harrington, P. M.; Kwon, B.-M. *J. Am. Chem. Soc.* **1992**, *114*,

1086. g) Buszek, K. R.; Jeong, Y. *Syn. Commun.* **1994**, *24*, 2461. h) Iida, K.; Hirama, M. *J. Am. Chem. Soc.* **1994**, *116*, 10310. i) Iida, K.; Hirama, M. *J. Am. Chem. Soc.* **1995**, *117*, 8875. j) Myers, A. G.; Hammond, M.; Wu, Y. S.; Xiang, J. N.; Harrington, P. M.; Kuo, E. Y. *J. Am. Chem. Soc.* **1996**, *118*, 10006. k) Sato, I.; Akahori, Y.; Iida, K.; Hirama, M. *Tetrahedron Lett.* **1996**, *37*, 5135. l) Kawata, S.; Yoshimura, F.; Irie, J.; Ehara, H.; Hirama, M. *Synlett* **1997**, 250.
- 3) a) Maier, M.; Brandstetter, T. *Tetrahedron Lett.* **1991**, *32*, 3679. b) Myers, A. G.; Dragovich, P. S. *J. Am. Chem. Soc.* **1992**, *114*, 5859. c) Audrain, H.; Skrydstrup, T.; Ulibarri, G.; Grierson, D. S. *Synlett* **1993**, 20. d) Audrain, H.; Skrydstrup, T.; Ulibarri, G.; Riche, C.; Chiaroni, A.; Grierson, D.S. *Tetrahedron* **1994**, *50*, 1469. e) Nuss, J. M.; Murphy, M. M. *Tetrahedron Lett.* **1994**, *35*, 37. f) Bunnage, M. E.; Nicolaou, K. C. *Angew. Chem. Int. Ed. Engl.* **1996**, *35*, 1110. g) Shibuya, M.; Wakayama, M.; Naoe, Y.; Kawakami, T.; Ishigaki, K.; Nemoto, H.; Shimizu, H.; Nagao, Y. *Tetrahedron Lett.* **1996**, *37*, 865.
- 4) a) Hirama, M.; Gomibuti, T.; Fujiwara, K. *J. Am. Chem. Soc.* **1991**, *113*, 9851. b) Nicolaou, K. C.; Schreiner, E. P.; Iwabuchi, Y.; Suzuki, T. *Angew. Chem. Int. Ed. Engl.* **1992**, *31*, 340. c) Boger, D. L.; Zhou, J. *J. Org. Chem.* **1993**, *58*, 3018. d) Tokuda, M.; Fujiwara, K.; Gomibuti, T.; Hirama, M.; Uesugi, M.; Sugiura, Y. *Tetrahedron Lett.* **1993**, *34*, 669. e) Semmelhack, M. F.; Gallapher, J. J.; Ding, W.-D.; Krishnamurthy, G.; Babune, R.; Ellestad, G. A. *J. Org. Chem.* **1994**, *59*, 4357. f) Wittman, M. D.; Kadow, J. F.; Langeley, R. D.; Yyas, D. M.; Rose, W. C.; Solomon, W.; Zein, N. *Bioorg. Med. Chem. Lett.* **1995**, *5*, 1049. g) Toshima, K.; Ohta, T.; Ohashi, A.; Nakamura, T.; Nakata, M.; Tatsuta, K.; Matsumura, S. *J. Am. Chem. Soc.* **1995**, *117*, 4822.
- 5) Takahashi, T.; Tanaka, H.; Yamada, H.; Matsumoto, T.; Sugiura, Y. *Angew. Chem. Int. Ed. Engl.* **1997**, *36*, 1524.
- 6) Doi, T.; Takahashi, T. *J. Org. Chem.* **1991**, *56*, 3465.
- 7) Takahashi, T.; Tanaka, H.; Hirai, Y.; Doi, T.; Yamada, H.; Shiraki, T.; Sugiura, Y. *Angew. Chem. Int. Ed. Engl.* **1993**, *32*, 1657.
- 8) NMR data of **2** (270 MHz, CDCl₃ : CD₃OD = 5 : 1): δ = 7.96–8.02 (m, 2H; Ph), 7.56–7.64 (m, 1H; Ph), 7.43–7.50 (m, 2H; Ph), 5.94–5.80 (m, 3H; vinyl, C5 and C12), 5.49–5.42 (m, 1H; vinyl), 5.37–5.35 (m, 1H; C11), 5.26–5.22 (m, 1H; vinyl), 4.36–4.31 (m, 2H; C10 and anomer), 4.13–4.00 (m, 2H; C4 and OCH₂RS-1H), 3.91–3.76 (m, 3H; OCH₂RS-1H and sugar-2H), 3.48–3.33 (m, 6H; SCH₂COOR and sugar-4H), 2.91–2.87 (m, 2H; SCH₂RO), 2.70–2.63 (m, 1H; C8-1H), 2.56–2.49 (m, 1H; C8-1H), 0.90 (m, 9H; (CH₃)₃CSi), 0.14–0.10 (m, 6H; CH₃Si); IR (KBr): ν = 3362, 2922, 1717, 1262, 1069, 838, 779, 709 cm⁻¹; FAB-MS:*m/z*: 745 [M⁺+H], 743 [M⁺-H].
- 9) NMR data of **1** (270 MHz, CD₃OD): δ = 8.47 (d, 1H, *J* = 8.6 Hz; Ph), 7.95 (d, 1H, *J* = 8.9 Hz; Ph), 7.50–7.82 (m, 6H; Ph), 7.37 (brdd, 1H, *J* = 6.9, 6.9 Hz; Ph), 7.15 (d, 1H, *J* = 8.9 Hz; Ph), 6.30–6.25 (m, 1H; C12), 6.03–6.10 (m, 1H; C11), 5.78–5.95 (m, 3H; vinyl), 5.40 (brd, 1H, *J* = 17.1 Hz; vinyl), 5.21 (brd, 1H, *J* = 9.9 Hz; vinyl), 4.27 (brd, 1H, *J* = 7.6 Hz; anomer), 3.97–4.13 (m, 2H; C4 and OCH₂RS), 3.75–3.95 (m, 5H; RCOOCH₃, sugar-1H and OCH₂RS-1H), 3.60–3.70 (m, 1H; sugar-1H), 3.47–3.57 (m, 2H; SCH₂COOR), 3.42(s, 3H; ROCH₃), 3.23–3.37 (m, 3H; sugar-3H), 3.13–3.25 (m, 1H; sugar-1H), 2.80–2.95 (m, 4H; SCH₂RO and C8); IR (CDCl₃): ν = 2922, 2848, 1727, 1644, 1579, 1461, 1265, 1202, 1132, 1069, 830, 753 cm⁻¹; FAB-MS:*m/z*: 911 [M⁺+K], 895 [M⁺+Na], 871 [M⁺-H].